

Article

# A Proposed Mechanistic and Translational Framework for Assessing Reproductive Toxicity of Hexavalent Chromium (Cr(VI))

Asmaa M. Neamah<sup>\*1</sup>

**Citation:** Neamah A. M. A Proposed Mechanistic and Translational Framework for Assessing Reproductive Toxicity of Hexavalent Chromium (Cr(VI)). American Journal of Biology and Natural Sciences 2026, 3(3), 23-32.

Received: 19<sup>th</sup> Dec 2025

Revised: 27<sup>th</sup> Jan 2026

Accepted: 18<sup>th</sup> Feb 2026

Published: 01<sup>st</sup> Mar 2026



**Copyright:** © 2026 by the authors. Submitted for open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>)

1. Department of Environment, College of Science, University of Al-Qadisiyah, Al Diwaniyah, Iraq

**Abstract:** Cr(VI) is a highly mobile and a very powerful environmental toxicant, which is well-known to be Multi-organ toxic and globally spread. There are growing reasons to suspect that Cr(VI) has profound effects on the reproductive health of men and women, but existing evaluations are still disjointed on human epidemiological observations, animal research and mechanistic research. This research paper suggests a mechanistic-translational paradigm to be used in standardized assessment of Cr(VI)-induced reproductive toxicity. According to this work, regular marshaling of evidence provided by occupational and environmental human exposure research with controlled in vivo and in vitro research study yields consistent reproductive endpoints, dose-response relationships, and common pathways. In all the exposure models, Cr(VI) is always linked with poor sperm quality, hormonal interference, ovarian impairment, adverse pregnancy, and developmental toxicity. These actions all converge mechanistically upon oxidative stress generation, DNA damage, mitochondrial dysfunction, apoptosis, and interference with endocrine systems. Notably, Human and animal data can be aligned to allow the development of causal plausibility as well as enhance the reliability of risk assessment. To expand on this cross-model synthesis, we suggest a systematic evaluation plan, which is a combination of molecular biomarkers, functional reproductive outcomes, and developmental outcomes into a single evaluation plan. The framework will help to improve detection at the early stage, the decision-making process by regulators, and facilitate the elaboration of specific prevention and remediation plans. The offered model offers a scientifically sound basis in the process of developing an environmental reproductive risk assessment of Cr(VI) and the corresponding metal toxicants.

**Keywords:** Cr (VI), Environmental Pollutants, Reproductive Toxicity, Oxidative Stress, Fertility Impairment.

## Introduction

Cr(VI) is a very mobile, water soluble, and highly oxidizing environmental pollutant whose systemic toxicity has been established. Its continued presence in industrial effluents, groundwater systems and in the workplace has caused chronic human and ecological exposures across the globe.

The anthropogenic sources are majorly electroplating, stainless-steel welding, tanning of leather, production of pigments, cement production and chemical processing by use of chromates. Cr(VI) is a compound that is highly bioavailable and is able to penetrate cell membranes through sulfate transporters thus readily accumulating in the biosphere and is reduced intracellularly to form reactive intermediates and reactive oxygen species (ROS). All these biochemical characteristics put Cr(VI) in a high-priority category as an environmental toxicant with multi-organ effects [1], [2], [3].

Although carcinogenicity and pulmonary effects of Cr(VI) exposure have been well studied there has been growing evidence on the high reproductive toxicity of the agent in both sexes. Human epidemiological research indicates a relationship between occupational exposure and poor semen parameters, hormonal disproportion, menstrual abnormalities, and poor pregnancy outcome. Similar results in experimental animal models show dose-dependent changes in spermatogenesis, ovarian follicular wellbeing, endocrine indication, and developing embryonic development. Molecularly, exposure to Cr(VI) causes oxidative stress, breaks in DNA strands, mitochondrial dysfunction, apoptotic signaling as well as epigenetic changes that all damage reproductive competence [3], [4], [5].

However, the existing evaluation of the Cr(VI)-induced reproductive toxicity remains incoherent even with the growing evidence. Human information, animal studies and mechanistic investigations are often considered independently without the chance of the translation to affect risk determination and prejudice the risk assessment dependability. Moreover, there is a lack of uniformity in the reporting of reproductive endpoints between studies and no standardized strategies of assessment based on biomarkers. This loophole minimizes the ability to translate mechanistic results into regulatory frameworks and those related to the regulation of health to the people [6], [7].

To overcome these shortcomings, this paper offers a mechanistic-translational model of assessing the reproductive toxicity triggered by Cr(VI). Instead of being a descriptive review, this work has combined epidemiological, *in vivo*, and mechanistic evidence to determine converging pathways, everywhere reproducible endpoints, and dose-response trends in biological models. Through systematic mapping of the molecular processes to functional reproductive outcomes, we have a systematic assessment strategy that correlates cellular processes with clinical and developmental outcomes [6], [8], [9].

Methodologically, the framework is built by methodically integrating evidence of peer-reviewed studies conducted on humans and animals on the reproductive parameters, endocrine functionality, and aspects of developmental toxicity. A comparative study was conducted to find out the consistency in the exposure models, mechanistic plausibility, and translational relevance. The emphasis was given on the biomarkers of oxidative stress, mitochondrial dysfunction, apoptotic processes, endocrine dysregulation and dose-response characterization, which represent the mechanistic structure of the proposed model of evaluation [4], [10].

Through this integrative endeavor, the present study will contribute to scientific comprehension of Cr(VI) reproductive toxicology and will provide a consistent platform on how the future experimentation work, biomarkers validation, and regulatory risk assessment would be made to be performed.

The purpose of this paper is to critically review the reproductive toxicity of hexavalent chromium by reviewing the human, animal, and *in vitro* studies of the subject. Specifically, these are the objectives to:

- The reproductive risks of exposure to Cr(VI) in men, such as its effects on sperm quality, hormonal levels, and testicular activity.
- Assess the reproductive impacts of Cr(VI) on females, specifically, on ovarian functionality, oocyte quality, pregnancy, and fetus development.
- Research on the molecular and cellular pathways of reproductive toxicity caused by Cr(VI), such as oxidative stress, DNA damage, apoptosis, and endocrine disruption.
- Compare findings in epidemiological, *in vivo*, and *in vitro* to determine similarities and differences among exposure models.
- Find the existing gaps in the research and provide directions on how future research can be improved to learn and avoid Cr(VI)-related reproductive toxicity.

## Materials and Methods

The paper is a summary of the accessible human and animal data that tends to evaluate the reproductive toxicity of Cr ( VI ).

### 2.1 Conceptual Design of the Framework

This study was proposed as an evidence-integration study that was aimed at producing a mechanistic-translational model to determine the toxicity-induced by Cr(VI) on reproduction. The methodology was focused on a systematic mapping of convergent biological endpoints in human epidemiology research, experimental animal models and mechanistic research, as opposed to a conventional review of the literature. This was to identify reproducible trends, similar molecular pathways and dose-response associations that would be brought into a standardized evaluation plan.

### 2.2 Evidence Identification and Selection Strategy

The peer-reviewed studies were found with the help of the targeted search in the large scientific databases, including PubMed, Scopus, and Web of Science. The search strategy involved the combination of the following keywords:

hexavalent chromium OR Cr(VI) and reproductive toxicity or fertility or spermatogenesis or ovarian toxicity or developmental toxicity or pregnancy outcome or oxidative stress or mitochondrial dysfunction.

Published studies in the English language were taken into consideration. Human and animal studies were incorporated in case they gave a measure of reproductive outcomes or mechanistic biomarkers following exposure to Cr(VI).

#### 1. Inclusion criteria:

- Occupational or environmental Cr(VI) exposure.
- Quantifiable reproductive outcomes (e.g., sperm parameters, hormone levels, estrous cyclicity, fetal growth).
- Mechanistic evaluation (e.g., ROS, apoptosis, DNA damage, mitochondrial dysfunction).
- Dose-response assessment where available.

#### 2. Exclusion criteria:

- Studies lacking reproductive endpoints.
- Reports without clear exposure characterization.
- Studies focusing exclusively on non-reproductive organ systems.
- This strategy ensured that only evidence directly relevant to reproductive assessment was integrated into the framework.

### 2.3 Translational Evidence Mapping

Selected studies were categorized into three analytical tiers:

- Human epidemiological evidence.
- In vivo animal models.
- Molecular and cellular mechanistic studies.
- Endpoints were grouped into functional reproductive outcomes (e.g., sperm motility, ovarian reserve, fetal weight), endocrine parameters, and mechanistic biomarkers.

Cross-model comparison was performed to determine:

- Consistency of reproductive alterations.
- Dose-dependent trends.
- Biological plausibility of mechanisms.
- Translational alignment between species.

The comparison of the mapping enabled the recognition of mechanistic nodes in both human and animal data, which reinforced the use of cause and effect interpretation.

### 2.4 Mechanistic Integration Model

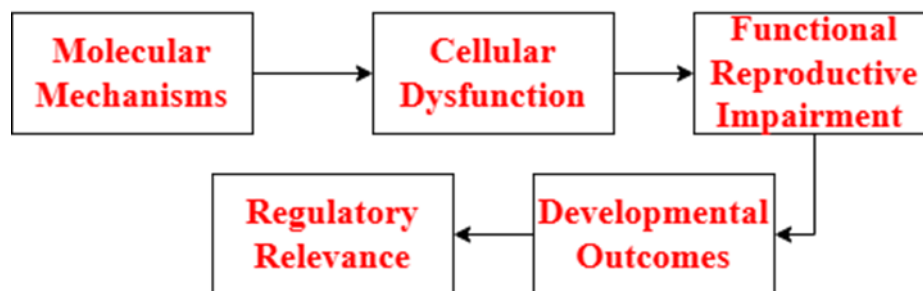
Endpoints that were mechanistic were prioritized in cases where they were supported by evidence at several levels of biology. Specific attention was given to:

- Oxidative stress indicators (MDA, SOD, ROS generation).
- DNA integrity markers (fragmentation, strand breaks).
- Apoptotic signaling pathways (caspase activation).
- Mitochondrial dysfunction markers.
- Endocrine disruption parameters.

It was these components that were incorporated into a hierarchical model of molecular disturbances to cellular damage, organ-level dysfunction, and clinical reproductive outcomes.

## 2.5 Framework Construction

The last framework was built by aligning the molecular mechanisms with cell dysfunction, pathology at the organ-level, and functional outcomes of the reproductive system, which is schematically shown in Figure 1.



**Figure 1.** The final framework constructed by aligning sequential biological events across hierarchical levels.

This gradual framework will offer a standard method by which the experimental design and risk assessment are to be applied in the future.

## Results and Discussion

### 3.1 Epidemiological Evidence in Humans

#### 3.1.1 Male Reproductive Effects

Occupational exposure to hexavalent chromium (Cr(VI)) has been linked to detrimental effects on the male reproductive system. Human research has also noted severe depletion of sperm count and sperm motility in workers who received Cr(VI) in contrast to untreated control group and signifies dysfunctional spermatogenesis as well as semen quality. Also, evidence of increased percentages of morphologically abnormal sperm exist with exposed individuals and this may indicate the possibility of genotoxic effect on spermatozoa [11], [12].

These findings are also supported by experimental studies in rodents; subacute exposure of Cr(VI) compounds led to a decrease in sperm count, reduction in motility, aberrant forms in sperm, and changes in reproductive hormones (e.g. increased FSH and reduced testosterone) and histological changes to testicular tissue [12], [13].

Mechanistically, Cr(VI)-impaired oxidative stress leads to DNA damage and dysfunction of the cell in spermatogenic cells, diminishing male fertility even more [11].

#### 3.1.2 Female Reproductive Effects

Exposure to hexavalent chromium (Cr(VI)) in women both occupational and environmental has been linked to negative reproductive effects. It is postulated through epidemiological and experimental evidence that exposure can result in menstrual abnormalities, hormonal derangements, poor quality oocytes and infertility [11]. Mechanistic plausibility of human reproductive toxicity An animal study suggests that Cr(VI) is capable of causing oxidative stress in ovarian tissue, oocyte maturation disruption and risk of premature embryonic loss, which is considered to be mechanistically plausible

[14]. Moreover, there is evidence of environmental exposure to the impact of Cr(VI)-contaminated environments on poor pregnancy outcomes with preterm delivery and low birth weight although in some cases human data are limited by confounding variables like co-exposure to other metals [15].

### 3.1.3 Developmental and Pregnancy Outcomes

Exposure of the maternal system to hexavalent chromium (Cr(VI)) has been linked to poor development and pregnancy. It has been experimentally shown that prenatal exposure to Cr(VI) can result in fetal growth retardation, elevated rates of resorption, and structural defects to a great extent through oxidative stress and placental dysfunction. Animal models show that Cr(VI) is easily absorbed across the placental barrier and causes DNA damage and apoptosis in embryonic tissues thus impairment of normal fetal development [11].

Associations have also been reported between high environmental exposure to chromium and high risk of stillbirth, preterm birth and low birth weight in epidemiological studies in localities with contamination of chromium compounds but human data is still limited partially because of other confounding environmental factors [16]. In general, existing toxicological and epidemiological data indicate that Cr(VI) exposure could have a dramatic impact on the normal course of pregnancy and development of the fetus [15].

## 3.2 Evidence from Animal Studies

### 3.2.1 Male Reproductive Toxicity

The strong mechanistic evidence is given on the basis of experimental animal studies that support the Cr(VI)-induced male reproductive toxicity. It has been demonstrated that subchronic exposure to hexavalent chromium in rats causes extensive sperm count and motility losses, abnormal sperm morphology, damage to seminiferous tubules, Leydig cell damage, and testosterone reduction. Histopathological results indicate that the tumors of the testes exhibit disorganization of the testicular tissue and cellular damage caused by oxidative stress [17], [18]. These dose effects reinforce the causal relationship between the exposure of Cr(VI) and male reproduction dysfunction.

### 3.2.2 Female Reproductive Toxicity

Exposure to Cr(VI) in the female rodents has been linked to follicular atresia, reduced oocyte count, estrous cycle disruption, hormonal imbalance, and augmented granulosa cell division. The evidence of experiments suggests that oxidative stress is a key factor in the dysfunction of the ovary, and the weakening of antioxidant defense mechanisms in the ovary has been observed in the tissues of the ovary [19]. There is also a decreased implantation, higher rates of fetal resorptions and placental abnormalities caused by gestational exposure, which prove multi-level reproductive toxicity of females [15].

### 3.2.3 Developmental and Embryotoxic Effects

Animal models of prenatal exposure to Cr(VI) have been associated with growth retardation of the fetus, skeletal deformity, neurodevelopmental defects, higher rates of embryo malformations, and teratogenesis in a dose-dependent effect. Research shows that Cr(VI) has the capacity to bypass the placental barrier, cause oxidative DNA damages, and disable placental functioning, thus affecting the ability to transfer nutrients into the fetus [20]. The results indicate the embryotoxic and developmental effect of Cr(VI).

## 3.3 Dose-Response Relationship

The result of animal toxicological studies is always consistent, and they show a definite dose-response relationship between Cr(VI) exposure and reproductive toxicity. Concentration and time of exposure are associated with higher levels of oxidative stress, endocrine disturbance and histopathological damage of reproductive tissues. Environmentally relevant doses have been sufficient to induce measurable reproductive impairment in experimental models [18], [20]. Toxicological evaluations have identified No-Observed-Adverse-Effect Levels (NOAELs) and Lowest-Observed-Adverse-Effect Levels (LOAELs) for chromium compounds in reproductive studies, although chronic exposure often exceeds LOAEL thresholds, resulting in pronounced reproductive toxicity [20].

Cr(VI) exerts multicomponent and overlapping molecular action on the reproductive toxicity. Cr(VI) gets in cells through sulphate transporters and is reduced stepwisely to Cr(III) producing reactive oxygen species (ROS) which damage gametophilic integrity, mitochondrial activity and redox balance. Damage to the DNA in the sperm and oocytes interferes with cell division resulting in chromatin abnormalities and development defects. The interference with endocrine functions changes the levels of steroidogenic enzymes and hormones, disrupting the processes of spermatogenesis and estrous cyclicity, in addition to the implantation. Mitochondrial dysfunction also diminishes the availability of energy, lowering the sperm motility and oocytes quality. Also, the combination of apoptosis in seminiferous tubules and granulosa cells as well as alterations in the epigenome, including the use of DNA methylation and the imbalanced expression of microRNAs, leads to long-term reproductive dysfunction. Table 1 and Table 2 show, respectively, the human and animal reproductive outcomes of exposure to Cr(VI) and the most important endpoints, all the observed changes, and the referencing sources.

**Table 1.** Summary of Human Reproductive Effects of Cr(VI) Exposure.

Gender	Endpoint	Observed Effect	Reference
Male	Testosterone	Altered levels	[1]
Male	DNA damage	Increased fragmentation	[1]
Female	Ovarian function	Oocyte developmental disruption	[11]
Female	Pregnancy outcomes	Low birth weight, fetal growth restriction	[21]
Developmental	Fetal growth	Growth impairment (SGA risk)	[21]

**Table 2.** Summary of Animal Reproductive Effects of Cr(VI) Exposure.

Species	Gender	Endpoint	Observed Effect	Reference
Rat	Male	Sperm count	Decreased	[22]
Rat	Male	Motility	Reduced	[22]
Rat	Male	Testicular histology	Seminiferous tubule degeneration	[22]
Rat	Male	Testosterone	Decreased	[22]
Rat	Female	Oocyte development	Chromosomal abnormalities	[11]
Rat	Female	Follicular structure	Atresia	[11]
Rat	Developmental	Fetal growth	Reduced fetal weight	[15]
Rat	Developmental	Placental function	Dysfunction	[15]

The summary presented in Tables 1 and 2 demonstrates strong concordance between human epidemiological findings and experimental animal studies regarding Cr(VI)-induced reproductive toxicity. In men, work-related exposure has been linked throughout with a reduction in the count of sperm and motor capacity, changing degrees of testosterone, and augmented DNA harm [2]. These changes are in line with the mechanistic rodent studies that indicated degeneration of seminiferous tubules and oxidative stress-induced testicular damage [22].

In women, it has been shown that Cr(VI) interferes with the functioning of the ovary and development of the fetus in humans and animals. The results of the experiments indicate the presence of oocyte damage and chromosomal instability caused by oxidative stress and epidemiological evidence indicates that it is related to fetal growth restriction and adverse pregnancy outcomes [11], [21]. Animal studies which show the low fetal weight and placental dysfunction following maternal exposure are also supportive of developmental toxicity [15].

In general, the convergent effect of the mechanistic processes across the species justifies the fact that Cr(VI) has a reproductive toxic effect on the organism by causing oxidative stress, DNA damage, endocrine disruption, mitochondrial dysfunction, and apoptosis. These multi-level effects support the necessity of severe occupational and environmental exposure control.

### Proposed Mechanistic–Translational Assessment Framework

We grow on the integrative evidence that has been acquired with the help of the human epidemiological evidence, animal-based experimentation and mechanistic investigations to propose a stringent mechanistic-translational judgment framework to assess the reproductive toxicity of Cr (VI). The framework will determine a systematic connection between the dysfunction and molecular perturbations on an organ level and their effects on the reproductive outcomes as perceived by the clinical setting. Compared to classical descriptive assessments, the proposed model integrates the bio-events at hierarchical degrees of organization and matches them with the quantifiable biomarkers and risk assessment suitability. The model is formulated into five developmental levels of biology:

1. Molecular Initiation Events.
2. Cellular Dysfunction.
3. Tissue and Organ-Level Alterations.
4. Functional Reproductive Outcomes.
5. Developmental and Population-Level Implications.

All levels have convergent species and exposure-based evidence.

#### 1. Tier 1 – Molecular Initiation Events

Cr(VI) enters cells via sulfate transport pathways and undergoes intracellular reduction to Cr(III), generating reactive oxygen species (ROS). This initiates:

- Oxidative stress.
- DNA strand breaks.
- Epigenetic alterations.
- Mitochondrial respiratory chain impairment.

These initial molecular processes are the major toxicity provokers.

#### 2. Tier 2 – Cellular Dysfunction

Oxidative imbalance and mitochondrial dysfunction cause cellular levels to result in:

- Apoptosis activation (caspase pathways).
- Granulosa cell degeneration.
- Leydig cell dysfunction.
- Impaired ATP production.

This step has a direct negative effect on the gamete viability and endocrine signaling.

#### 3. Tier 3 – Tissue and Organ-Level Alterations

Persistent cell damage develops into:

- Seminiferous tubule degeneration.
- Ovarian follicular atresia.
- Placental insufficiency.
- Hormonal dysregulation.

These changes are also constantly seen in animal models and contribute to the impaired reproductive functions in humans.

#### 4. Tier 4 – Functional Reproductive Outcomes

Pathology at the organ level occurs as quantifiable level of reproduction.:

- Decreased sperm count and motility.
- Abnormal sperm morphology.
- Reduced testosterone levels.
- Estrous cycle disruption.
- Reduced fertility and implantation success.

These are endpoint translational markers involving the relationship between mechanistic toxicity and clinical relevance.

## 5. Tier 5 – Developmental and Population-Level Impact

In high levels of exposure or in chronic instances:

- Fetal growth retardation.
- Low birth weight.
- Spontaneous abortion.
- Embryotoxicity.

It is this level of the implications of the intergenerational effects of exposure to Cr(VI).

The structured components of the proposed mechanistic–translational assessment model, including hierarchical biological tiers, key mechanistic events, measurable biomarkers, and their translational relevance, are summarized in Table 3.

**Table 3.** Proposed Mechanistic–Translational Assessment Model for Cr(VI)-Induced Reproductive Toxicity.

Tier	Biological Level	Key Events	Measurable Biomarkers	Translational Outcome	Risk Assessment Relevance
Tier 1	Molecular	ROS generation, DNA damage, epigenetic changes	MDA, SOD, DNA fragmentation, 8-OHdG	Cellular stress initiation	Early toxicity indicator
Tier 2	Cellular	Apoptosis, mitochondrial dysfunction	Caspase-3, ATP depletion, mitochondrial membrane potential	Gamete impairment	Mechanistic validation
Tier 3	Tissue/Organ	Testicular degeneration, ovarian atresia, endocrine disruption	Histopathology, hormone assays	Organ dysfunction	Reproductive hazard identification
Tier 4	Functional	Reduced sperm quality, hormonal imbalance, and fertility decline	Semen analysis, estrous cycle assessment	Clinical reproductive impairment	Translational endpoint
Tier 5	Developmental	Fetal growth restriction, embryotoxicity	Birth weight, fetal viability, and placental weight	Intergenerational risk	Regulatory significance

## Conclusion

Cr(VI) is one of the significant reproductive risks that are associated with toxicity of various organs and species and are of great environmental and human health challenges at the global level. There must be an interdisciplinary approach to effective management that combines regulatory control, monitoring of the environment, and technological intervention. The most effective remediation method is the reduction of Cr(VI) to Cr(III) which is less toxic, but it should be further perfected to make it suitable in large scales. Reduction of human and ecological exposure is reliant on the stringent surveillance, implementation of normative standards and embracing of green manufacturing activities. More sensitive detection strategies, investigation of greener remedial measures and the assessment of

long term reproductive outcomes especially in the vulnerable and high-risk groups should be a priority area of the research in future.

## REFERENCES

- [1] M. Costa, "Toxicity and carcinogenicity of chromium compounds in humans," *Critical Reviews in Toxicology*, vol. 27, no. 5, pp. 431–442, 1997.
- [2] L. Wuri, J. A. Arosh, J. Z. Wu, and S. K. Banu, "Exposure to hexavalent chromium causes infertility by disrupting cytoskeletal machinery and mitochondrial function of the metaphase II oocytes in superovulated rats," *Toxicology Reports*, vol. 9, pp. 219–229, 2022, doi: 10.1016/j.toxrep.2022.02.002.
- [3] S. Kumar, N. G. Sathwara, A. K. Gautam, K. Agarwal, B. Shah, P. K. Kulkarni, K. Patel, A. Patel, L. M. Dave, D. J. Parikh, and H. N. Saiyed, "Semen quality of industrial workers occupationally exposed to chromium," *Journal of Occupational Health*, vol. 47, no. 5, pp. 424–430, 2005, doi: 10.1539/joh.47.424.
- [4] W. Zheng et al., "In utero exposure to hexavalent chromium disrupts rat fetal testis development," *Toxicology Letters*, vol. 299, pp. 201–209, 2018.
- [5] L. Wuri et al., "Exposure to hexavalent chromium causes infertility by disrupting cytoskeletal machinery and mitochondrial function," *Toxicology Reports*, vol. 9, pp. 219–229, 2022.
- [6] V. Singh, N. Singh, M. Verma, R. Kamal, and S. K. Banu, "Hexavalent chromium-induced oxidative stress and the protective role of antioxidants against cellular toxicity," *Antioxidants*, vol. 11, no. 12, Art. no. 2375, 2022, doi: 10.3390/antiox11122375.
- [7] U.S. Environmental Protection Agency, *IRIS Toxicological Review of Hexavalent Chromium (Cr(VI)) (CASRN 18540-29-9)*, Washington, DC, USA: U.S. EPA, 2022. Available: <https://www.ncbi.nlm.nih.gov/books/NBK611281/>
- [8] J.-J. Ding, C. Jiao, Y.-L. Qi, et al., "New insights into the reverse of chromium-induced reprotoxicity of pregnant mice by melatonin," *Ecotoxicology and Environmental Safety*, vol. 238, Art. no. 113608, 2022, doi: 10.1016/j.ecoenv.2022.113608.
- [9] E. Sazakli, "Human health effects of oral exposure to chromium: A systematic review of the epidemiological evidence," *International Journal of Environmental Research and Public Health*, vol. 21, no. 4, Art. no. 406, 2024, doi: 10.3390/ijerph21040406.
- [10] X. Xu, Y. Li, Z. Wang, et al., "Hexavalent chromium exposure induces reproductive toxicity via oxidative stress pathways," *Regulatory Toxicology and Pharmacology*, vol. 123, Art. no. 104966, 2021, doi: 10.1016/j.yrtph.2021.104966.
- [11] L. Wuri, R. C. Burghardt, J. Arosh, C. R. Long, and S. K. Banu, "Hexavalent chromium disrupts oocyte development in rats by elevating oxidative stress, DNA double-strand breaks, microtubule disruption, and aberrant segregation of chromosomes," *Int. J. Mol. Sci.*, vol. 24, no. 12, p. 10003, Jun. 2023. DOI: 10.3390/ijms241210003.
- [12] N. Marouani, O. Tebourbi, S. Mahjoub, M. T. Yacoubi, M. Sakly, M. Benkhalifa, and K. B. Rhouma, "Effects of hexavalent chromium on reproductive functions of male adult rats," *Reproductive Biology*, vol. 12, no. 2, pp. 119–133, Jul. 2012, DOI: 10.1016/S1642-431X(12)60081-3.
- [13] H. Li et al., "Effect of Cr(VI) exposure on sperm quality: human and animal studies," *Ann. Occup. Hyg.*, vol. 45, no. 7, pp. 505–511, Oct. 2001, DOI: 10.1093/annhyg/45.7.505.
- [14] N. Marouani, O. Tebourbi, S. Mahjoub, M. T. Yacoubi, M. Sakly, and K. B. Rhouma, "Effects of hexavalent chromium on reproductive functions of male adult rats," *Reproductive Biology*, vol. 12, no. 2, pp. 119–133, 2012. doi: 10.1016/S1642-431X(12)60081-3.
- [15] Agency for Toxic Substances and Disease Registry (ATSDR), *Toxicological Profile for Chromium*. Atlanta, GA, USA: U.S. Department of Health and Human Services, 2012. [Online]. Available: <https://www.atsdr.cdc.gov/toxprofiles/tp7.pdf>.
- [16] Y. Fang, Q. Shen, K. Yang, M. Wang, W. Wang, J. Lv, M. Fang, M. Nian, Y. Huang, Z. Huang, L. Cui, D. Xu, and Y. Fan, "Effects of exposure to chromium during pregnancy on fetal growth and

- a possible sex-dependent response: Results of a cross-sectional study," *Biological Trace Element Research*, vol. 204, no. 1, pp. 74–84, Jan. 2026, doi: 10.1007/s12011-025-04664-4.
- [17] N. Marouani, O. Tebourbi, S. Mahjoub, M. T. Yacoubi, M. Sakly, and K. B. Rhouma, "Effects of hexavalent chromium on reproductive functions of male adult rats," *Reproductive Biology*, vol. 12, no. 2, pp. 119–133, 2012. doi: 10.1016/S1642-431X(12)60081-3.
- [18] M. Costa and C. B. Klein, "Toxicity and carcinogenicity of chromium compounds in humans," *Critical Reviews in Toxicology*, vol. 36, no. 2, pp. 155–163, 2006. doi: 10.1080/10408440600681422.
- [19] L. Wuri, R. C. Burghardt, J. Arosh, C. R. Long, and S. K. Banu, "Hexavalent chromium disrupts oocyte development in rats by elevating oxidative stress, DNA double-strand breaks, microtubule disruption, and aberrant segregation of chromosomes," *International Journal of Molecular Sciences*, vol. 24, no. 12, p. 10003, 2023. doi: 10.3390/ijms241210003.
- [20] Y. Fang et al., "Effects of exposure to chromium during pregnancy on fetal growth and a possible sex-dependent response," *Biological Trace Element Research*, vol. 204, no. 1, pp. 74–84, 2026. doi: 10.1007/s12011-025-04664-4.
- [21] Y. Fang et al., "Effects of exposure to chromium during pregnancy on fetal growth," *Biological Trace Element Research*, vol. 204, no. 1, pp. 74–84, 2026. doi: 10.1007/s12011-025-04664-4.
- [22] N. Marouani et al., "Effects of hexavalent chromium on reproductive functions of male adult rats," *Reproductive Biology*, vol. 12, no. 2, pp. 119–133, 2012. doi: 10.1016/S1642-431X(12)60081-3.